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COX, LOX and platelet aggregation inhibitory properties of Lauraceae neolignans

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ABSTRACT

The anti-inflammatory potential of 26 neolignans (14 of the bicyclooctane-type and 12 of the benzofuran-type), isolated from three Lauraceae species (*Pleurothyrium cinereum*, *Ocotea macrophylla* and *Nectandra amazonum*), was evaluated in vitro through inhibition of COX-1, COX-2, 5-LOX and agonist-induced aggregation of rabbit platelets. Benzofuran neolignans were found to be selective COX-2 inhibitors, whereas bicyclooctane neolignans inhibit selectively the PAF-action as well as COX-1 and 5-LOX. The neolignan 9-nor-7,8-dehydro-isolicarin B **15** and cinerin C **7** were found to be the most potent COX-2 inhibitor and PAF-antagonist, respectively. Nectamazin C **10** exhibited dual 5-LOX/COX-2 inhibition.

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Cyclooxygenase (COX) and lipoxygenase (LOX) are the two major enzyme families that catalyze the rate-limiting step in the formation of prostanoids, prostaglandins (PGs), and thromboxane A₂ (TxA₂) by the COX pathway, and leukotrienes (LTs) by the LOX pathway, whose products are significant mediators of pain, fever and inflammation.¹ Inflammatory effects result both from direct actions of PGs and LTs (on the microvasculature, on nociceptive afferents and on temperature-regulating centers in the hypothalamus) and, indirectly, by synergy with other inflammatory mediators including bradykinin, histamine, activated component and platelet activating factor (PAF).² There are three isoforms of COX, namely a constitutive form (COX-1) that is present in many tissues such as platelets, stomach, lungs, kidneys etc, an inducible form (COX-2) that is expressed during inflammation as a result of stimulation by cytokines, nitric oxide and growth factors, and a splice variant of COX-1 (COX-3).^{2,3} A great interest in developing inhibitors specific for COX-2 had increased in the last years, expecting that it will lack the adverse effects caused by the inhibition of COX-1 enzyme. However, considering the pro-inflammatory properties of prostanoids, a current interest had been focused for dual 5-LOX and COX-2 inhibitors, which had emerged as a rational advance for the design of efficacious anti-inflammatory agents.^{1,4}

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It has been demonstrated that platelets play an important role in acute inflammation.⁵ They accumulate and respond to injury by releasing important mediators such as 5-HT, PGs, PAF and hydro-lases.⁶ Furthermore, TxA₂, formed by platelets has been reported to be potent constrictor of blood vessels and an aggregator of platelets,⁷ which are activated by several chemical agents such as arachidonic acid (AA), adenosine diphosphate (ADP), collagen, epinephrine, platelet activating factor (PAF), among others.⁸ PAF was discovered to be a lipid mediator of hypersensitivity and inflammation.⁹ Several studies have implicated PAF in such diseases as asthma, hypertension, cardiac anaphylaxis and arthritis as well as its clinical benefits on these cases.¹⁰

As part of our search for bioactive neolignans from Lauraceae plants, a phytochemical exploration was carried out on the leaves of *Pleurothyrium cinereum*, *Ocotea macrophylla* and *Nectandra amazonum*, afforded 26 neolignans, 14 related to the bicyclooctane-type (cinerins A-D¹¹ **1–4**, ocophyllols A-C¹² **5–7**, nectamazins A-C¹³ **8–10**, kadsurenin C¹³ **11**, 4'-oxo-macrophyllin B¹⁴ **12**, macrophyllin B¹⁴ **13** and 2'-epi-guianin¹² **14**), and 12 related to the 8,5',7,0,4'-connected (9'-nor-7,8-dehydro-isolicarin B¹⁵ **15**, (–)- and (+)-licarin B^{12,14} **16–17**, (–)- and (+)-licarin A^{13,14} **18–19**, (+)-mirandatin A¹⁴ **20**, ocophyllals A-B¹² **21–22**, (+)-acuminatin¹³ **23**, (+)-denudatin B¹³ **24**, (+)-kadsurenone **25** and lilifloll A **26**). The structures and the absolute configuration (Figs. 1 and 2) of the former compounds were determined by extensive spectroscopic analyses, whose isolation and structural elucidation is discussed in previous papers.^{11–15}

Several Lauraceae plants have exhibited anti-inflammatory properties.^{16–19} The Lauraceae chemistry is recognized to comprise

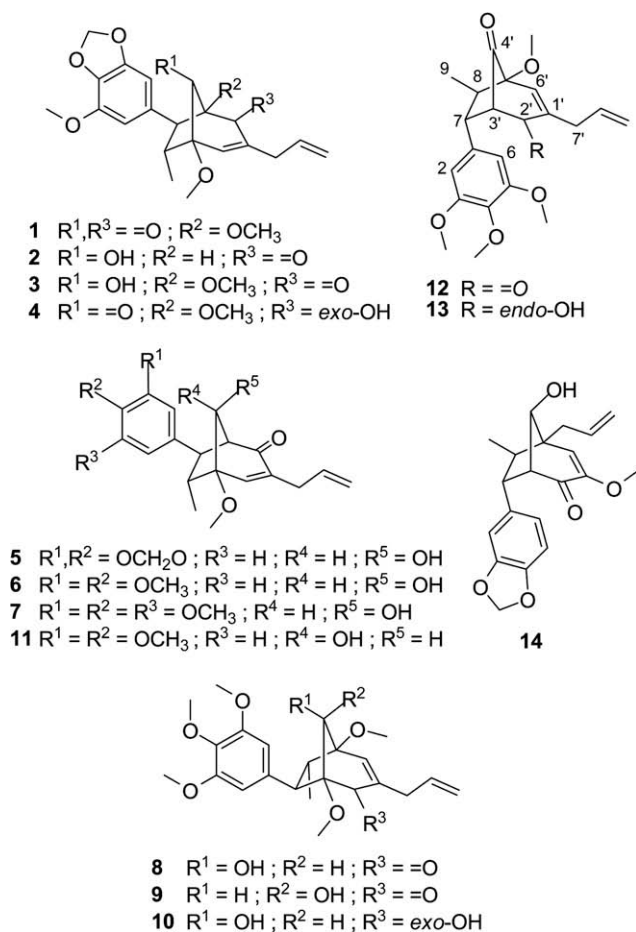


Figure 1. Structures of the bicyclo[3.2.1]octane neolignans.

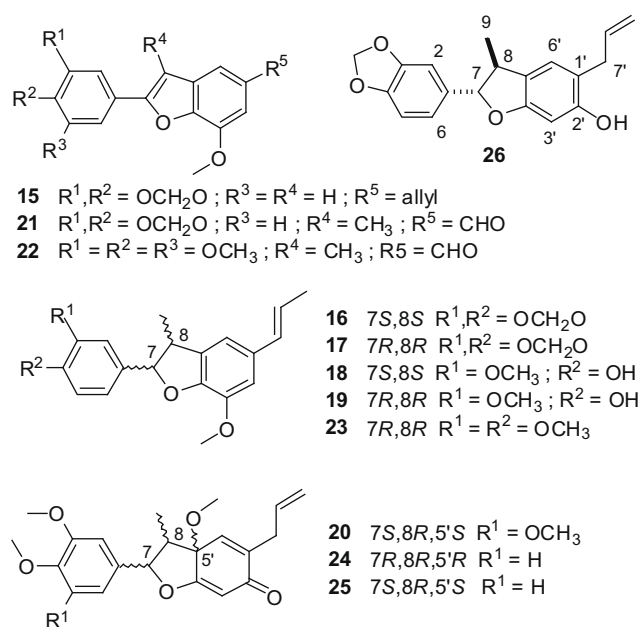


Figure 2. Structures of the 8.5',7.0.4'-connected neolignans.

neolignans, mostly related to the 8.5',7.0.4'-connected class, which have shown in vitro anti-inflammatory activity through suppression of tumor necrosis factor (TNF)- α and nitric oxide (NO) production,²⁰ and inhibitory activity against the two isozymes of COX.²¹

On the basis of these facts and in order to establish the potential as anti-inflammatory agents of the isolated compounds, an in vitro screening for the capability to inhibit the cyclooxygenase (COX) isozymes (COX-1 and COX-2) and 5-lipoxygenase (5-LOX) were accomplished, using a COX-(ovine) and a 5-LOX-(potato) inhibitor screening kits, respectively, following the reported methodologies.²²

The in vitro abilities (IC_{50} values, μM) of the isolated compounds (**1–26**) to inhibit the isozymes COX-1 and COX-2 were determined in the COX-catalyzed transformation of AA into PGH_2 , which is then reduced to $\text{PGF}_{2\alpha}$ and detected by an enzyme immunoassay (EIA). Although only a work reported the COX-1 and COX-2 inhibition of four bicyclooctane diastereomers (isolated from the stem bark of *Ocotea bullata*, a medicinal plant from southern Africa), which exhibited no inhibitory effect,²³ the neolignans **1–14** showed COX-inhibition at different levels (Tables 1 and 2). The macrophyllin-type¹⁴ neolignans **2–3, 5–7, 11, 14** as well as the guianin-type¹⁴ neolignan **14** exhibited selectivity toward COX-1 inhibition (IC_{50} values 18.2–94.9 μM range).

Compounds **2** and **11** (which have identical bicyclooctane moiety, being solely differentiated by the aryl-substitution) have similar IC_{50} values (32.5 and 38.7 μM , respectively), suggesting that the aryl-substitution was not a significant structural condition in the COX-1 inhibition for this type of neolignans, although 3,4,5-trioxyphenyl neolignans showed slightly higher inhibition than 3,4-dioxyphenyl neolignans, as shown for compounds **5–7**. However, the activity was notably influenced by the configuration of the bicyclooctane moiety. This statement is supported on comparing the activity for the C-4'-epimers **8** and **9**, which possess a C-4' hydroxyl group placed in opposite orientation. Compound **8** (with OH group oriented toward aryl group) exhibited a higher IC_{50} value to that of **9** (with OH group oriented toward enone). Similar result was observed between compounds **2** and **6**.

Although the neolignans with a carbonyl group at C-4' (like **1, 4, 12–13**) instead C-4' OH group showed weaker COX-1 inhibitory activity, compounds having a C-2' hydroxyl group (like **4, 10** and **13**) showed selective COX-2 inhibition, which is supported on comparing the IC_{50} values of the compounds **12** and **13**, whose only difference is the functional group at C-2' (C-2' carbonyl group in **12** and C-2' *endo*-hydroxyl group in **13**). Compounds **10** and **14** were found to be the most potent COX-1 and COX-2 inhibitors (IC_{50} 18.2 and 6.83 μM , respectively), among the bicyclo[3.2.1]octane neolignans.

Table 1
COX-1, COX-2 and 5-LOX enzyme inhibition of bicyclo[3.2.1]octane neolignans **1–14**

| Compds | IC_{50}^a (μM) | | SI ^b | IC_{50}^a (μM) |
|--------------|--------------------------------------|-------|-----------------|--------------------------------------|
| | COX-1 | COX-2 | | 5-LOX |
| 1 | 165 | 288 | 0.573 | 45.6 |
| 2 | 32.5 | 215 | 0.151 | 146 |
| 3 | 28.3 | 356 | 0.0795 | 136 |
| 4 | 188 | 92.1 | 2.04 | 8.84 |
| 5 | 88.6 | 312 | 0.284 | 256 |
| 6 | 94.9 | 288 | 0.330 | 189 |
| 7 | 72.1 | 245 | 0.294 | 176 |
| 8 | 72.2 | 255 | 0.283 | 156 |
| 9 | 152 | 569 | 0.266 | 42.4 |
| 10 | 74.6 | 6.83 | 10.9 | 12.8 |
| 11 | 38.7 | 312 | 0.124 | 117 |
| 12 | 356 | 844 | 0.422 | 92.5 |
| 13 | 216 | 65.4 | 3.30 | 18.9 |
| 14 | 18.2 | 326 | 0.0558 | 113 |
| Celecoxib | 8.32 | 0.113 | 73.4 | 11.3 |
| Aspirin | 0.411 | 2.53 | 0.162 | — |
| Caffeic acid | — | — | — | 3.74 |

^a Values are means of two experiments, standard deviation from the mean is <10% of the mean value.

^b In vitro COX-2 selectivity index (IC_{50} COX-1/ IC_{50} COX-2).

Table 2
COX-1, COX-2 and 5-LOX enzyme inhibition of 8,5',7,0,4'-connected neolignans **15–26**

| Compds | IC ₅₀ ^a (μM) | | SI ^b | IC ₅₀ ^a (μM) 5-LOX |
|--------------|------------------------------------|-------|-----------------|---|
| | COX-1 | COX-2 | | |
| 15 | 349 | 3.32 | 106 | 329 |
| 16 | 366 | 25.6 | 14.3 | 426 |
| 17 | 289 | 52.1 | 5.55 | 157 |
| 18 | 312 | 32.1 | 9.73 | 35.5 |
| 19 | 245 | 66.4 | 3.69 | 12.5 |
| 20 | 77.9 | 126 | 0.620 | 13.2 |
| 21 | 123 | 16.8 | 7.31 | 436 |
| 22 | 102 | 12.7 | 8.02 | 412 |
| 23 | 223 | 28.7 | 7.75 | 146 |
| 24 | 88.6 | 446 | 0.199 | 15.6 |
| 25 | 25.6 | 246 | 0.104 | 12.6 |
| 26 | 179 | 98.7 | 1.81 | 10.2 |
| Celecoxib | 8.32 | 0.113 | 73.4 | 11.3 |
| Aspirin | 0.411 | 2.53 | 0.162 | — |
| Caffeic acid | — | — | — | 3.74 |

^a Values are means of two experiments, standard deviation from the mean is <10% of the mean value.

^b In vitro COX-2 selectivity index (IC₅₀ COX-1/IC₅₀ COX-2).

In the case of the COX inhibitory activity displayed by the 8,5',7,0,4'-connected neolignans, it was found that the neolignans having a dihydrobenzofuran core (**16–19**, **23**, **26**) selectively inhibited the COX-2 enzyme at moderate level (IC₅₀ 25.6–98.7 μM range). The levorotatory dihydrobenzofuran neolignans **16** and **18** were found to be slightly more active than the dextrorotatory compounds **17** and **19**.

The neolignans having a benzofuran moiety (**15**, **21–22**) also exhibited COX-2 selectivity, affording the best results (IC₅₀ 3.32–16.8 μM range). Thus, compound **15** was the most active COX-2 inhibitor (IC₅₀ 3.32 μM), among neolignans **1–26**. Although **15** was much weaker COX-2 inhibitor than the selective COX-2 inhibitor celecoxib (1.5% of the celecoxib's potency), the COX-2 selectivity index of **15** was found to be higher (COX-2 SI = 106), suggesting further studies toward structural optimization for increasing the activity, perhaps by preparation of derivatives of **15** containing the COX-2 pharmacophores methanesulfonyl (MeSO₂) or sulfonamide (H₂NSO₂) as substitutions.^{4,22} In contrast, the neolignans with dihydrobenzofuran-(2H)-one moiety (**20**, **24–25**) inhibited selectively the COX-1 action.

In vitro 5-LOX enzyme inhibition studies indicate that compounds **4**, **10**, **13**, **19–20**, **24–26** had comparable 5-LOX inhibition activity than celecoxib, but lesser than caffeic acid. Interestingly, compound **10** had a dual 5-LOX/COX-2 inhibition (COX-2 IC₅₀ 6.83 μM; 5-LOX IC₅₀ 12.8 μM), which actually is the main intention for the development of new anti-inflammatory agents. This dual inhibition of **10** might be likely due to the ability for binding to, or chelate iron present in the 5-LOX enzyme, by their groups at C-2' and C-3', whose orientation places them spatially nearby. Thus, **10** is a good candidate to be used as dual inhibitor in further anti-inflammatory studies. As expected, neolignans having a free phenolic OH group (**18**, **19**, **26**) showed 5-LOX inhibition, but their COX-inhibition was no significant. The dihydrobenzofuran-(2H)-one neolignans (**20**, **24–25**) were found to be equipotent (IC₅₀ values in the 12.6–15.6 μM range). However, further structure–activity studies and biological analyses are required to clarify the underlying mechanism and to draw unambiguous conclusions for the COX-inhibition for these classes of neolignans.

The capability to inhibit the agonist-induced platelet aggregation had been previously evaluated for lignans and neolignans induced by agonists such as platelet activating factor (PAF), arachidonic acid (AA), adenosine 5'-diphosphate (ADP), adrenaline, among others.^{24–26} In addition, some 8,5',7,0,4'-connected and

bicyclo[3.2.1]octane neolignans isolated from Chinese medicinal plants were found to be potent PAF-antagonists in the ³H-PAF receptor binding assay.²⁷ In previous papers, we report the inhibition of PAF-induced platelet aggregation of neolignans **1–11**.^{11–13} However, in order to establish selectivity toward the common agonist identified in platelet aggregation, it is also described herein a comparison of the abilities to inhibit the platelet aggregation induced by PAF (7.20 nM), AA (100 μM), and ADP (4.00 μM) for the neolignans **1–26** (Table 3), following the reported methodology.¹⁰

Clear trends were observed in this screening. ADP-induced platelet aggregation was no significantly inhibited by the evaluated compounds **1–26**. Macrophyllin-type bicyclooctane neolignans **1–13** showed a noticeable selectivity toward PAF-inhibition on comparing the IC₅₀ values when the other two agonists were used, whilst compound **14** (a guianin-type neolignan) was found to be a non-selective inhibitor. Macrophyllin-type neolignans having a C-4' hydroxyl group (like **2–3**, **5–11**, **14**) exhibited higher inhibition toward PAF (IC₅₀ in the 1.09–3.78 μM range values) than neolignans with C-4' keto group (IC₅₀ in the 6.63–16.8 μM range values), while neolignans having a C-2' hydroxyl group (like **4**, **10** and **13**) showed higher AA-antagonism. In addition, if the C-4' hydroxyl group was oriented toward aryl group (like **2–3**, **8**) the PAF-inhibition was slightly increased (IC₅₀ in the 1.09–1.37 μM range values).

Kadsurenone **25**, a recognized PAF-antagonist from the Chinese medicinal plant *Piper futokadsura*,^{28,29} was also isolated from *N. amazonum*. In addition to the excellent inhibition to PAF, **25** showed activity for AA and ADP at different levels. Interestingly, the activity of mirandin A **20**, which solely differs from **25** by an aromatic O-methyl group, was lower to that of **25**. Furthermore, the configuration was found to be an important factor for inhibition of platelet aggregation, since the diastereomer denudatin B **24** exhibited activity significantly lower to that of **20**. The benzofuran-type 8,5',7,0,4'-connected neolignans **15–19**, **21–23** and **26** exhibited no significant activity for platelet aggregation. Additionally to kadsurenone **25**, cinerins B–C **2–3**, and nectamazin A **8** were

Table 3
Inhibitory effects^a of neolignans **1–26** on the aggregation of rabbit platelets induced by PAF, AA and ADP

| Compds | PAF (7.20 nM) ^a | AA (100 μM) ^a | ADP (4.00 μM) ^a |
|--------------|----------------------------|--------------------------|----------------------------|
| 1 | 16.8 ± 1.6 | 75.6 ± 1.6 | 88.9 ± 4.5 |
| 2 | 1.51 ± 0.35 | 42.8 ± 1.5 | >999 |
| 3 | 1.09 ± 0.23 | 85.8 ± 1.1 | >999 |
| 4 | 6.63 ± 0.75 | 25.6 ± 0.9 | 356 ± 33 |
| 5 | 3.08 ± 0.15 | 45.6 ± 2.3 | 636 ± 25 |
| 6 | 3.03 ± 0.45 | 38.8 ± 3.2 | 725 ± 58 |
| 7 | 2.34 ± 0.66 | 48.6 ± 1.8 | >999 |
| 8 | 1.37 ± 0.45 | 68.7 ± 1.0 | >999 |
| 9 | 1.72 ± 0.47 | 43.4 ± 0.8 | >999 |
| 10 | 3.78 ± 0.12 | 21.6 ± 0.5 | >999 |
| 11 | 2.33 ± 0.23 | 54.6 ± 1.2 | >999 |
| 12 | 7.47 ± 0.89 | 78.9 ± 0.9 | 521 ± 63 |
| 13 | 6.79 ± 0.78 | 23.2 ± 2.1 | 121 ± 38 |
| 14 | 1.61 ± 0.15 | 2.62 ± 1.03 | 23.1 ± 1.1 |
| 15 | 460 ± 13 | 426 ± 15 | >999 |
| 16 | 488 ± 22 | 290 ± 22 | 54.6 ± 2.3 |
| 17 | 565 ± 25 | 326 ± 32 | 56.7 ± 3.4 |
| 18 | >999 | 489 ± 28 | 112 ± 12 |
| 19 | >999 | 556 ± 45 | 165 ± 25 |
| 20 | 3.28 ± 0.31 | 38.7 ± 1.7 | 121 ± 18 |
| 21 | >999 | 568 ± 35 | 102 ± 11 |
| 22 | 847 ± 32 | 425 ± 56 | 98.3 ± 2.1 |
| 23 | 65.3 ± 9.8 | 222 ± 33 | 75.6 ± 3.2 |
| 24 | 75.6 ± 1.3 | 58.3 ± 2.8 | 148 ± 16 |
| 25 | 0.153 ± 0.018 | 5.61 ± 0.98 | 38.1 ± 2.8 |
| 26 | 18.4 ± 3.3 | 145 ± 19 | 124 ± 27 |
| Aspirin | 10.3 ± 1.8 | 5.33 ± 0.45 | 545 ± 36 |
| Gingkolide B | 0.925 ± 0.097 | 75.6 ± 1.0 | >999 |

^a The data were expressed as means 95% confidence intervals of four rabbits.

the most potent PAF-antagonists. Compound **14** were the most potent inhibitor for platelet aggregation induced by PAF, AA and ADP.

In summary, a set of 26 naturally-occurring neolignans, related to the bicyclo[3.2.1]octane and 8.5',7.0.4'-connected classes, were evaluated in order to establish their ability to inhibit COX-1, COX-2, 5-LOX and platelet aggregation induced by PAF, AA and ADP. In the case of bicyclooctanes, the macrophyllin-type neolignans **2–3**, **5–7**, **11**, **14** as well as the guianin-type neolignan **14** exhibited selectivity toward COX-1 inhibition, whilst compounds **4**, **10** and **13** showed selective COX-2 inhibition, being the activity notably influenced by the configuration of the bicyclic part. In addition, it was found that the neolignans having a dihydrobenzofuran core (**16–19**, **23**, **26**) selectively inhibited the COX-2 isozyme and the levorotatory were found to be slightly more active than dextrorotatory dihydrobenzofurans. Compound **15** was the most active COX-2 inhibitor (whose IS was higher than celecoxib), and compound **10** had a dual 5-LOX/COX-2 inhibition. Macrophyllin-type bicyclooctane neolignans **1–13** exhibited a noticeable selectivity toward PAF-inhibition. These results imply that the neolignans isolated from *P. cinereum*, *O. macrophylla* and *N. amazonum* might be beneficial in the treatment of inflammatory and vascular diseases.

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References and notes

- Charlier, C.; Michaux, C. *Eur. J. Med. Chem.* **2003**, *38*, 645.
- Simmons, D. L.; Botting, R. M.; Hla, T. *Pharmacol. Rev.* **2004**, *56*, 387.
- Chandrasekharan, N. V.; Dai, H.; Roos, K. L. T.; Evanson, N. K.; Tomsik, J.; Elton, T. S.; Simmons, D. L. *Proc. Natl. Acad. Sci. U.S.A.* **2002**, *99*, 13926.
- Chowdhury, M. A.; Abdellatif, K. R. A.; Dong, Y.; Rahman, M.; Das, D.; Suresh, M. R.; Knaus, E. E. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 584.
- Vincent, J. E.; Bonta, I. L.; Zijlstra, F. J. *Agents Actions* **1978**, *8*, 291.
- Page, C. P.; Paul, W.; Morley, J. *Int. Arch. Allergy Appl. Immunol.* **1984**, *74*, 347.
- Lefer, A. M.; Messenger, M.; Okamatsu, S. *Naunyn. Schmiedeberg's Arch. Pharmacol.* **1982**, *321*, 130.
- Siess, W. *Physiol. Rev.* **1989**, *69*, 58.
- Levi, R.; Burke, J. A.; Guo, Z. G.; Hattori, Y.; Hoppens, C. M.; McManus, L. M.; Hanahan, D. J.; Pinckard, R. N. *Circ. Res.* **1984**, *54*, 117.
- Koch, E. *Phytomedicine* **2005**, *12*, 10.
- Coy, E. D.; Cuca, L. E.; Sefkow, M. J. *Nat. Prod.* **2009**, *72*, 1245.
- Coy, E. D.; Cuca, L. E.; Sefkow, M. *Phytochemistry* **2009**, *70*, 1309.
- Coy, E. D.; Cuca, L. E. *Chem. Pharm. Bull.* **2009**, *57*, 639.
- Coy, E. D.; Cuca, L. E. *Biochem. Syst. Ecol.* **2008**, *36*, 674.
- Coy, E. D.; Cuca, L. E. *Rev. Colomb. Quim.* **2008**, *37*, 127.
- Phan, M. G.; Phan, T. S.; Matsunami, K.; Otsuka, H. *Chem. Pharm. Bull.* **2006**, *54*, 380.
- Zschocke, S.; Drewes, S. E.; Paulus, K.; Bauer, R.; van Staden, J. *J. Ethnopharmacol.* **2000**, *71*, 219.
- Moreno, S. R. F.; Arnobio, A.; De Carvalho, J. J.; Nascimento, A. L.; Timoteo, M. O.; Olej, B.; Rocha, E. K.; Pereira, M.; Bernardo-Filho, M.; Caldas, L. Q. *A. Biol. Res.* **2007**, *40*, 131.
- da Silva Filho, A. A.; Albuquerque, S.; de Silva, M. L. A.; Eberlin, M. N.; Tomazela, D. M.; Bastos, J. K. J. *Nat. Prod.* **2004**, *67*, 42.
- Cho, J. Y.; Baik, K. U.; Yoo, E. S.; Yoshikawa, K.; Park, M. H. *J. Nat. Prod.* **2000**, *63*, 1205.
- Su, B. N.; Cuendet, M.; Hawthorne, M. E.; Kardono, L. B. S.; Rswan, S.; Fong, H. H. S.; Mahta, R. G.; Pezzuto, J. M.; Kinghorn, A. D. *J. Nat. Prod.* **2002**, *65*, 163.
- Chowdhury, M. A.; Abdellatif, K. R. A.; Dong, Y.; Das, D.; Suresh, M. R.; Knaus, E. E. *Bioorg. Med. Chem. Lett.* **2008**, *18*, 6138.
- Zschocke, S.; van Staden, J.; Paulus, K.; Bauer, R.; Horn, M. M.; Munro, O. Q.; Brown, N. J.; Drewes, S. E. *Phytochemistry* **2000**, *54*, 591.
- Wang, B. G.; Hong, X.; Li, L.; Zhou, J.; Hao, X. J. *Planta Med.* **2001**, *66*, 511.
- Chen, Y. C.; Liao, C. H.; Chen, I. S. *Phytochemistry* **2007**, *68*, 2101.
- Yang, Y. P.; Cheng, M. J.; Teng, C. M.; Chang, Y. L.; Tsai, I. L.; Chen, I. S. *Phytochemistry* **2002**, *61*, 567.
- Han, G. *Prog. Nat. Sci.* **1995**, *5*, 299.
- Chang, M. N.; Han, G. Q.; Arison, B. H.; Springer, J. P.; Hwang, S. B.; Shen, T. Y. *Phytochemistry* **1985**, *24*, 2079.
- Hwang, S. B.; Lam, M. H.; Shen, T. Y. *Adv. Inflammation Res.* **1986**, *11*, 83.